

AMCoR

Asahikawa Medical University Repository <http://amcor.asahikawa-med.ac.jp/>

Clinical Journal of Gastroenterology (2010) 3(3):117–127.

Novel perspectives in probiotic treatment : the efficacy and unveiled mechanisms of the physiological functions

Fujiya, Mikihiro ; Kohgo, Yutaka

Novel perspectives in probiotic treatment: The efficacy and unveiled mechanisms of the physiological functions

Mikihiro Fujiya, Yutaka Kohgo

Division of Gastroenterology and Hematology/Oncology, Department of Medicine,
Asahikawa Medical College

Running title

Novel perspectives in probiotic treatment

Key words

Probiotics, inflammatory bowel disease, irritable bowel syndrome, intestinal infection, intestinal barrier function, immunity, cell membrane transporters, novel organic cation transporter 2, Toll-like receptors, competence and sporulation factor, heat shock proteins

Correspondence

To whom correspondence should be addressed:

Yutaka Kohgo, MD, Ph.D.

Division of Gastroenterology and Hematology/Oncology, Department of Medicine,

Asahikawa Medical College, Asahikawa, Japan

2-1 Midorigaoka-higashi, Asahikawa, Hokkaido 078-8510, Japan

Tel: +81-166-68-2462

Fax: +81-166-68-2469

e-mail: yk1950@asahikawa-med.ac.jp

Abstract

Probiotics are defined as “live microorganisms which confer a health benefit on the host” when administered in adequate amounts, and have potential effects for maintaining intestinal development, nutrition, and treating intestinal inflammations, functional disorders, and other extra-intestinal diseases. Although the benefits of probiotics for human health were first noted over 100 years ago, the analysis of probiotic functions began in earnest only 20 years ago. Probiotics, such as some strains of *Lactobacillus*, *Bifidobacterium*, *Escherichia coli*, and *Bacillus subtilis*, inhibit the growth of pathogenic bacteria, induce competitive effects for the adherent of pathogenic bacteria and their toxins to intestinal epithelia, induce cytoprotective heat shock proteins, enhance the intestinal barrier function and modulate the host immune responses. The crosstalk between the host and the probiotics appears to be mediated by bacteria-derived effectors, which can be sensed with multiple systems, including the Toll-like receptors and cell membrane transporters. Future analyses will identify more probiotic-derived effectors, the recognition mechanisms of these effectors, and the subsequent changes of the intestinal epithelia and immune cells for each probiotic treatment. For clinical use, a procedure that objectively evaluates the ability of each probiotic effect will help establish a standard for choosing the most valuable strain and

its proper dose for each individual patient.

Introduction

The mammalian intestine continuously comes in contact with prokaryotes, which are indispensable partners for developing intestinal tissue and maintaining its homeostasis. The International Human Microbiome Project (1) (2) has provided a tremendous amount of information on commensal bacteria and has ushered in a new era for the field of gastroenterology. It is clear that more than 2,000 species of commensal bacterial organisms exist in our bodies, and the majority is located in the gut, but most of these bacteria are not pathogens (3). Recently, new ribosomal RNA- and whole genome-based technologies have highlighted that the populations of microbial species differ between individuals, regions of the gut axis (4), and different mucosal layers at a single anatomical site, thus illustrating the diverse and complicated role of “the partner” in our biological activities (5) (6).

Probiotics, which are defined as “live microorganisms which confer a health benefit on the host, when administered in adequate amounts” (7), have potential effects for human health as well as the treatment of intestinal disorders. Indeed, many etiological studies have proposed the beneficial effects of probiotics since 100 years ago (8) (9) (10). Several recent meta-analyses and systematic reviews suggest that probiotics are potentially useful preventive or therapeutic strategies for multiple gastrointestinal

diseases, while the data from these studies were based on various subjects using different genera, species, strains, and doses of probiotics (11) (12) (13) (14) (15) (16) (17) (18). However, data from probiotic treatments remain preliminary thus far, and are influenced by the diversity of the microbiota, diets, and genetic backgrounds of the individual subjects. It is crucial to elucidate the mechanisms of certain probiotic functions. Early mechanistic investigations involved the remodeling of microbial communities to suppress the growth and activity of pathogens, particularly in acute infections. Subsequently, probiotics were found to cause various effects, including the upregulation of anti-inflammatory factors, immunomodulation via the suppression of pro-inflammatory mediators, enhanced immunity, epithelial cell differentiation and proliferation, and promotion of intestinal barrier function. In the present review, the clinical and physiological effects of probiotics and novel perspectives in the mechanistic studies of the probiotic effects are discussed.

1. Clinical benefits of probiotics

There is increasing evidence that probiotics are required for the development of a healthy intestinal system, the promotion of host nutritional status, and the treatment and prevention of intestinal infections, and functional disorders of the gastrointestinal

tract.

1) Intestinal development

It is known that the intestinal microbiota are important for the development of the normal gut, particularly in maintaining normal immunological reactions and cytoprotective responses, and also in regulating apoptosis (19) (20) (21). Indeed, LGG decreases chemically induced apoptosis and increases the expression of genes involved in cytoprotective responses in the developing mouse small intestine (22). The administration of *Lactobacillus casei* DN-114001 appears to ameliorate the gut immune response through the stimulation of macrophages and dendritic cells (23). These studies demonstrate that the probiotics may thus play a significant role in intestinal development.

2) Nutrition

Intestinal microbiota are heavily involved in the regulation of human and other host nutritional status, including the promotion of polysaccharide digestion and nutrient uptake and the regulation of the energy balance by cooperation with intestinal epithelial cells (24) (25). Intestinal microbiota regulate energy balance by using and

storing the calories harvested from the host's diet (26) (27). Furthermore, a recent study has shown the deviations from a gut core microbiome, which shares the genes involved in various metabolic functions, to be associated with aberrant physiological states, such as obesity (28). Supportive evidence concerning the probiotic effects on the prevention and treatment of obesity has been recently addressed by clinical trials (29) (30).

3) Intestinal disorders

A) Infectious diseases

There are many studies which reveal the therapeutic effects of probiotic treatment for several types of the infectious enteritis (31) (32). Travelers' diarrhea is one of the common diseases among travelers to foreign countries, particularly in Africa and Southeast Asia. From the data of more than 10 randomized controlled studies, probiotic treatments have been shown to be safe and effective for the prevention of travelers' diarrhea (33) (34). However, this effect appears to be dependent on the population of each study, the type of probiotics used, the duration of treatment, the trip destination, and the compliance with treatment. Otherwise, the therapeutic and preventive effects of probiotics such as VSL#3, a mixture of 8 probiotic bacteria, and *Lactobacillus rhamnosus GG (LGG)* for rotavirus diarrhea have also been proposed in randomized

controlled studies (35) (36).

B) Inflammatory bowel diseases

Inflammatory bowel disease (IBD) is a chronic inflammatory condition of the gastrointestinal tract. The most common types of IBD are ulcerative colitis (UC) and Crohn's disease (CD), with the prevalence of IBD estimated to be 150,000 patients in Japan. Although the precise etiology of IBD is still obscure, the alterations in the intestinal microbiota are thought to play an important role in the pathogenesis of IBD. Many trials were therefore conducted to manipulate the intestinal microflora using probiotics, including VSL#3, LGG, and E. coli Nissle 1917, and several potential therapeutic applications for UC have been conducted for the induction of remission and the relief of clinical symptoms (37) (38) (39) (40), maintenance of the clinical remission (41) (42), and prevention of the onset of pouchitis (43) (44) (45) (46). Although controversial results have also been reported (47), the administration of probiotics appears to be a promising modality for the treatment of UC. Conversely, the potential use of probiotics in the prevention or treatment of CD remains unclear. Several clinical studies have been performed to analyze the effects of probiotics on inducing clinical remission or maintaining surgically-induced remission using LGG or Lactobacillus johnsonii LA1. While one study found a modest reduction in the recurrence rates of

patients after a surgical resection by supplementation with *Lactobacillus johnsonii* LA1, other studies revealed no benefit of probiotic administration for CD treatment (**Table 1**).

The reason that no benefits were obtained by the administration of probiotics may be linked to a poor understanding of both the mechanisms of probiotics and the pathogenesis of CD.

C) Irritable bowel disease

Irritable bowel disease (IBS) is a functional gastrointestinal disorder with low-grade inflammation and immune responses as well as changes in the fecal microflora (48), suggesting potential probiotic benefits for its treatment. Indeed, various probiotics including *Lactobacillus*, *Bifidobacterium*, and *Saccharomyces* are shown to ameliorate IBS symptoms such as a decrease in flatulence, abdominal pain, and bloating (53) (54) (55) (56) (57) (58). Zeng et al. proposed that the fermented milk containing *Streptococcus thermophilus*, *Lactobacillus bulgaricus*, *L. acidophilus*, and *B. longum* decreases small bowel permeability in patients with diarrhea-predominant IBS and improves mucosal barrier function for IBS patients (54). Other systematic reviews on basic and clinical studies have also emphasized the effectiveness of probiotics on IBS treatment (55) (56) (57) (58). Standardized species and strain selections and the optimal dose of probiotics for IBS therapy will likely be established for clinical applications in

future.

D) Necrotizing enterocolitis

Necrotizing enterocolitis (NEC) is an inflammatory necrosis of the intestine, typically observed in premature infants. A number of studies have shown that probiotic treatment reduces the incidence as well as the severity of NEC (59) (60). A meta-analysis demonstrated that probiotics supplementation, including Bifidobacteria (*B. infantis*, *B. bifidum*, and *B. breve*), Lactobacillia (LGG, *L. acidophilus*), *S. thermophilus* and *S. boulardii*, and *E. coli* Nissle exhibited beneficial effects for NEC treatment (61). However, in general, probiotic treatment is regarded to be less effective for reducing the risk of sepsis and the number of days on total parenteral nutrition (62) (63). The mechanisms responsible for probiotic effects in the treatment of NEC patients are still poorly understood.

E) Antibiotic-associated enteritis

Antibiotic-associated enteritis is frequently observed in patients administered antibiotics, which can lead to the suppression of the normal gut microflora, thus causing a change in the gut microflora and the overgrowth of pathogenic bacteria. Typical antibiotic-associated enteritis includes acute hemorrhagic enteritis and pseudomembranous enterocolitis. Various meta-analyses have shown that probiotics

successfully prevent antibiotic-associated enteritis (64) (65) (66). Probiotics aid the recovery of the gut microflora following antibiotic impairment. Antibiotics-associated enteritis is one of the diseases that benefit the most from probiotic treatment.

4) Extra-intestinal disorders

A) Helicobacter pylori infection

Recent clinical trials revealed the beneficial effects of probiotics with antibiotic treatment for the eradication of *Helicobacter pylori* (*H. pylori*) infection (67) (68) (69). Almost all of the studies utilized *Lactobacillae* for these treatments. The mechanisms of *H. pylori* infection eradication were mediated by the antibiotic functions of lactic acid, bacteriocins, or other active components, competition for *H. pylori* adhesion to the gastric epithelia, enhancement of mucosal barrier function, and the modification of inflammatory mediators (70) (71) (72) (73) (74) (75) (76) (77) (78) (79). Probiotics can also relieve the side effects for the eradication of *H. pylori* infection (80) (81) (82) (83) (84) (85) (86) (87). However, no evidence has yet shown that probiotic treatment directly eradicates *H. pylori*. The clinical indications of probiotics for the treatment and eradication of *H. pylori* infection still remain unclear.

B) HIV infection

It is known that early HIV replication and CD4⁺ cell destruction frequently occur in the gastrointestinal tract. HIV-associated enteropathy is one of the critical outcomes of AIDS. A pilot study proposed that yogurt containing *L. rhamnosus* GR-1 and *L. reuteri* RC-14 improved both diarrhea and CD4⁺ cell number in HIV-infected patients (88) (89). Furthermore, the combination of *L. rhamnosus* GR-1, *L. reuteri* RC-14, and antibiotics increase the cure rate of bacterial vaginosis (90), thus suggesting a beneficial probiotic effect in both the management of HIV enteropathy and in reduced disease progression.

B) Others

It has also been reported that probiotic treatment is useful for curing urinary tract infections (91) and atopic diseases (92).

2. Mechanisms of probiotic functions

As mentioned in the previous section, probiotics are gaining widespread inclusion as a new preventive or therapeutic strategy for multiple gastrointestinal and extra-gastrointestinal diseases. However, these data are collected from an epidemiological analysis with the empirical selections of probiotics. In order to objectively determine how to choose the suitable strain for each disease, the mechanisms of probiotic functions must be clarified. In this regard, the mechanistic

analyses of probiotic functions for pathogenic bacteria as well as the host intestine are now under investigation (**Figure 1**).

1) Growth inhibition of pathogenic bacteria

Probiotics exhibit direct antibacterial effects for pathogenic bacteria through the production of antibacterial substances, including bacteriocins and acid (93) (94). Gram-positive bacteria such as *Lactococcus lactis* produce small antimicrobial peptides, or lantibiotics (93) (95), which have been found to be an inhibitor for pathogenic bacteria by targeting the lipid II component of the bacterial cell wall (96). Bacteriocins are other small antimicrobial peptides produced by Lactobacilli including *L. plantarum*, *L. acidophilus* NCFM, *L. johnsonii* NCC 533, and *L. sakei* (97)(98)(99)(100). Bacteriocins exert an antimicrobial effect with a narrow spectrum and toxicity for Gram-positive bacteria such as *Streptococcus*, *Staphylococcus*, *Lactococcus*, *Listeria*, and *Mycobacteria*. Bacteriocin acts by forming pores in the cytoplasmic membrane and inhibiting the essential enzyme activities of sensitive bacteria.

Strains of Lactobacilli produce acetic, lactic, and propionic acids that lead to inhibited growth of a wide range of Gram-negative pathogens, such as *Salmonella enteric*, by lowering the local pH (101). Moreover, other unknown bactericidal substances derived from the Lactobacillus strain exhibit antibacterial effects in a

pH-dependent manner (101) (102).

2) Inhibition of the adherence of pathogenic bacteria or their toxin to intestinal epithelia

Probiotics, including several strains of Lactobacilli and Bifidobacteria, competitively decrease the adhesion of both pathogenic bacteria and their toxins to the intestinal epithelium (103) (104) (105) (106). Pathogenic bacteria are thought to bind to intestinal epithelia through the interaction between bacterial lectins and carbohydrate moieties of glycoconjugate receptors on the epithelial surface. Probiotics have lectin-like adhesion and proteinase-like components, which mediate the inhibitory effect of probiotics for the adherence of pathogenic bacteria to the intestinal epithelium (107)(108)(109). Probiotics also function to block bacterial enterotoxin binding. A recombinant *E. coli* generated to express the glycosyltransferase genes and produce chimeric lipopolysaccharide, which neutralizes enterotoxins, has been reported to reduce mortality by virulent *V. cholerae* infection in mice (110) (111).

3) Inductions of cytoprotective heat shock proteins

Heat shock proteins (Hsp) are essential for the maintenance of intestinal homeostasis, by protecting colonic epithelial from injury and foreign stress (112) (113). Hsps expression is changed in the intestinal epithelia of IBD patients, suggesting the

association of Hsps in the pathogenesis of IBD (114) (115). Tao et al. has shown that soluble factors in LGG-conditioned media induced Hsps in intestinal epithelial cells, which was mediated by the activation of the p38- and JNK/MAPK pathway (116). We have also demonstrated that *Bacillus subtilis* (*B. subtilis*) and its conditioned media induced Hsps in Caco-2/bbe cells and mouse intestine. In addition, we found that the *B. subtilis*-produced factor, competence and sporulation factor (CSF) mediates the Hsp induction and protective effect of *B. subtilis* against oxidant stress (117), thus illustrating that the probiotic effects, at least in part, can be mediated by the induction of the cytoprotective Hsps.

4) Enhancement of intestinal barrier function

The intestinal barrier function is an important mechanism for defending epithelial cells against the attack of foreign virulence. The impairment of the intestinal barrier function causes the disruption of the intestinal integrity. Probiotics including *E. coli* Nissle 1917, LGG, *L. casei* DN-114 001, VSL#3, *L. acidophilus*, and *Bacteroides tetraiotaomicron* augment the intestinal barrier function following pathogenic bacteria, hydrogen peroxide, or cytokine-induced disruption of epithelial tight junctions (118) (119) (120) (121) (122). This improvement in intestinal barrier function is mediated through the expression and translocation of zonula occludens (ZO)-2, protein kinase C

(PKC), and/or the activating mitogen-activated protein kinases (MAPKs). Our recent study also demonstrated the protective effect of *B. subtilis* and its soluble factor CSF against oxidant stress-induced intestinal injury by promoting epithelial barrier function (117).

5) Modulations of host immune responses

Probiotics have the potential to modulate both innate and adaptive immunity. Recent reports suggest that fecal sIgA is increased by *Bifidobacterium animalis* (123) as well as nonviable LGG, which augments interleukin (IL)-6 secretion (124). *Escherichia coli* Nissle 1917 and VSL#3 induce β -defensin (125) (126), which is a pivotal antibacterial peptide preventing bacterial adherence and invasion. *B. tetaitaomicron* also stimulates the release of an antimicrobial peptide, angiogenin 4 from Paneth cells, suggesting the significant role of probiotics in host innate immunity.

Probiotics also exert an inhibitory effect on the production of pro-inflammatory mediators. LGG or its conditioned media inhibits lipopolysaccharide- (LPS-) or *Helicobacter pylori*-stimulated TNF production by murine macrophages (127). *E. coli* Nissle 1917 suppresses TNF, IFN- γ , and IL-2 release, and increases IL-10 expression by T-cells (128). *L. casei* Shirota decreases IL-6 and IFN- γ production induced by LPS in peripheral blood mononuclear cells of normal and colitis mice (129). LGG and *L.*

rhamnosus GR-1 also induce granulocyte-colony stimulating factor (G-CSF) from macrophages, which inhibits *E. coli*- or LPS-induced TNF production (130). On the other hand, it is proposed that VSL#3, *E. coli* Nissle 1917, *Lactobacillus reuteri*, and *L. casei* suppress excess inflammation through inducing IL-10 (128) (131) (132) (133). Conversely, probiotics enhance the host immune response when the host suffers from pathogenic bacteria infection. It is reported that dendritic cells take up a polysaccharide (PSA) of *Bacteroides fragilis*, which induces the maturation of dendritic cells and the production of Th1-type cytokines (134). Therefore, probiotics regulate the host immune responses for each condition through the modulation of immune cells such as dendritic cells, and by inducing production of pro- and anti-inflammatory factors.

NF- κ B signaling is a pivotal pathway for epithelial interactions with immune cells. The excess activation of this pathway was believed to cause intestinal injury by inflammatory disorder. Probiotics, including *L. reuteri*, LGG, *B. infantis*, and *L. salivarius*, have been reported to downregulate the expression of NF- κ B-induced pro-inflammatory mediators such as IL-8 in intestinal epithelial cells (135) (136) (137) (138). *Bacteroides thetaiotaomicron* and *Enterococcus faecalis* inhibit NF- κ B function through the promotion of the nuclear export of the RelA subunit of NF- κ B by the activation of peroxisome proliferators activated receptor (PPAR)- γ (139) (140). Therefore,

probiotics may exhibit an inhibitory effect on NF- κ B signaling, and thereby regulate excess inflammation. However, a recent study has proposed that the deficiency of NF- κ B leads to the promotion of intestinal epithelial apoptosis, impairment of antimicrobial peptide expression, and subsequent chronic inflammation (141). It is still unclear whether probiotics affect the regulation of the NF- κ B-related pathways and subsequent intestinal inflammation.

3. How do we sense probiotics?

As mentioned above, probiotics exhibit competitive effects for pathogenic bacteria as well as beneficial effects for host intestinal tissues, thus indicating the significance of the crosstalk system between the host and the probiotics. Some probiotic supernatants exert the beneficial effects similarly as probiotics themselves. The bacterial components or soluble factors released by bacteria in the supernatants are sensed by intestinal epithelia by some mechanism.

Toll-like receptors (TLRs), which belong to the family of pattern recognition receptors (PRRs), are expressed in both intestinal epithelial and dendritic cells. TLRs are candidates that recognize the factors or bacterial components, including microorganism-associated molecular patterns (MAMPs), flagella, lipoteichoic acid,

peptidoglycan, lipopolysaccharide, and cell wall-associated polysaccharide (CPS), originated from probiotics. It has been proposed that the physiological effect of *E. coli* Nissle 1917 for improving the intestinal inflammation of the mouse enteritis model is diminished in TLR2⁻ or TLR4-deficient mice (142). Conversely, the VSL#3 function for relieving mice colitis has also been reported to decrease in TLR9-deficient mice, but not in TLR2⁻ or TLR4-knockout mice (143), thus suggesting that specific TLRs are involved in the probiotic functions. However, the target PRR of each probiotic is unidentified, and it is unclear how TLRs distinguish the probiotic-derived molecules from those of pathogenic bacteria.

Our recent study shows that the *B. subtilis*-derived peptide CSF is imported by the epithelial cell membrane transporter protein (117), a novel organic cation transporter 2 (OCTN2), whose gene polymorphism is susceptible to Crohn's disease (144). Furthermore, the protective effects of *B. subtilis* and its soluble factor CSF on the intestinal epithelia are reduced by inhibition of OCTN2 function (117), suggesting that OCTN2 transport is essential for the effects of *B. subtilis* and CSF. This suggests a novel mechanism for sensing probiotics through the uptake of the soluble factors produced by bacteria. Consequently, multiple systems may be involved in recognizing and discriminating bacteria, including commensals, pathogens, and probiotics (**Figure**

2).

4. Future outlook

While more than 100 years have elapsed since the probiotic benefits on human health were noted, the mechanistic analysis of probiotic functions has been given recognized only in the past 20 years, and many of the mechanisms of probiotic effects are still unclear. It should be established the standard for selecting appropriate strain for the treatment of individual disease through clarifying the physiological function and their mechanism of each probiotics. While controversial findings have also been reported, the proper strain and dose of probiotics are essential for treating intestinal disorders, even neoplasms (145) (146) (147) (148) (149) (150) (151) (152). The identification of effective molecules produced by probiotics, which mediate the probiotic functions, is essential for providing clear mechanisms. This enables the development of novel therapeutic strategies for intestinal disorders using bacteria-derived effectors. Furthermore, the procedure to objectively evaluate and quantify the ability of each probiotic effect will help to establish a standard for choosing the most valuable strain and its proper dose for individual cases.

References

1. Turnbaugh PJ, Ley RE, Hamady M, Fraser-Liggett CM, Knight R, Gordon JI. The human microbiome project. *Nature* 2007;449:804–810.
2. Pennisi E. Metagenomics. Massive microbial sequence project proposed. *Science* 2007;315:1781.
3. McFall-Ngai M. Adaptive immunity: care for the community. *Nature* 2007;445:153.
4. Eckburg PB, Bik EM, Bernstein CN, Purdom E, Dethlefsen L, Sargent M, et al. Diversity of the human intestinal microbial flora. *Science* 2005;308:1635–1638.
5. Zoetendal EG, von Wright A, Vilpponen-Salmela T, Ben-Amor K, Akkermans AD, de Vos WM. Mucosa-associated bacteria in the human gastrointestinal tract are uniformly distributed along the colon and differ from the community recovered from feces. *Appl Environ Microbiol* 2002;68:3401–3407.
6. Preidis GA, Versalovic J. Targeting the human microbiome with antibiotics, probiotics, and prebiotics: gastroenterology enters the metagenomics era. *Gastroenterology*. 2009 May;136(6):2015-31.
7. World Health Organization. Health and nutritional properties of probiotics in food including powder milk with live lactic acid bacteria, 2001. Available at: http://www.who.int/foodsafety/publications/fs_management/en/probiotics.pdf.

8. Fuller R. Probiotics in man and animals. *J Appl Bacteriol* 1989; 66:365–378.
9. Metchnikoff E. *The prolongation of life: optimistic studies*. New York, NY: Springer Publishing Company, Inc, 2004.
10. Neish AS. Microbes in gastrointestinal health and disease. *Gastroenterology*. 2009 Jan;136(1):65-80.
11. Huang JS, Bousvaros A, Lee JW, Diaz A, Davidson EJ. Efficacy of probiotic use in acute diarrhea in children: a meta-analysis. *Dig Dis Sci* 2002; 47:2625–2634.
12. Allen SJ, Okoko B, Martinez E, Gregorio G, Dans LF. Probiotics for treating infectious diarrhoea. *Cochrane Database Syst Rev* 2004: CD003048.
13. Johnston BC, Supina AL, Ospina M, Vohra S. Probiotics for the prevention of pediatric antibiotic-associated diarrhea. *Cochrane Database Syst Rev* 2007:CD004827.
14. D'Souza AL, Rajkumar C, Cooke J, Bulpitt CJ. Probiotics in prevention of antibiotic associated diarrhoea: meta-analysis. *BMJ* 2002; 324:1361.
15. Nikfar S, Rahimi R, Rahimi F, Derakhshani S, Abdollahi M. Efficacy of probiotics in irritable bowel syndrome: a meta-analysis of randomized, controlled trials. *Dis Colon Rectum* 2008;51:1775–1780.
16. McFarland LV, Dublin S. Meta-analysis of probiotics for the treatment of irritable

- bowel syndrome. *World J Gastroenterol* 2008;14:2650–2661.
17. Alfaleh K, Bassler D. Probiotics for prevention of necrotizing enterocolitis in preterm infants. *Cochrane Database Syst Rev* 2008:CD005496.
 18. Deshpande G, Rao S, Patole S. Probiotics for prevention of necrotising enterocolitis in preterm neonates with very low birth-weight: a systematic review of randomised controlled trials. *Lancet* 2007;369:1614–1620.
 19. Mazmanian SK, Liu CH, Tzianabos AO, Kasper DL. An immunomodulatory molecule of symbiotic bacteria directs maturation of the host immune system. *Cell* 2005; 122:107–118.
 20. Bouskra D, Brézillon C, Bérard M, Werts C, Varona R, Boneca IG, et al. Lymphoid tissue genesis induced by commensals through NOD1 regulates intestinal homeostasis. *Nature* 2008; 456:507–510.
 21. Wagner RD. Effects of microbiota on GI health: gnotobiotic research. *Adv Exp Med Biol* 2008; 635:41–56.
 22. Lin PW, Nasr TR, Berardinelli AJ, Kumar A, Neish AS. The probiotic *Lactobacillus* GG may augment intestinal host defense by regulating apoptosis and promoting cytoprotective responses in the developing murine gut. *Pediatr Res* 2008; 64:511–516.

23. de Moreno de LeBlanc A, Dogi CA, Galdeano CM, Carmuega E, Weill R, Perdigón G. Effect of the administration of a fermented milk containing *Lactobacillus casei* DN-114001 on intestinal microbiota and gut associated immune cells of nursing mice and after weaning until immune maturity. *BMC Immunol* 2008; 9:27.
24. Yan F, Polk DB. Commensal bacteria in the gut: learning who our friends are. *Curr Op Gastroenterol* 2004; 20:565–571.
25. Yan F, Polk DB. Probiotics: progress toward novel therapies for intestinal diseases. *Current Opinion in Gastroenterology* 2010, 26:95–101.
26. Ley RE, Turnbaugh PJ, Klein S, Gordon JI. Microbial ecology: human gut microbes associated with obesity. *Nature* 2006; 444:1022–1023.
27. Turnbaugh PJ, Ley RE, Mahowald MA, Magrini V, Mardis ER, Gordon JI. An obesity-associated gut microbiome with increased capacity for energy harvest. *Nature* 2006; 444:1027–1031.
28. Turnbaugh PJ, Hamady M, Yatsunenko T, Cantarel BL, Duncan A, Ley RE, et al. A core gut microbiome in obese and lean twins. *Nature* 2009; 457:480–484.
29. Luoto R, Kalliomäki M, Laitinen K, Isolauri E. The impact of perinatal probiotic intervention on the development of overweight and obesity: follow-up study from birth to 10 years. *Int J Obes (Lond)*. 2010 Mar 16. [Epub ahead of print]

30. Kadooka Y, Sato M, Imaizumi K, Ogawa A, Ikuyama K, Akai Y, et al. Regulation of abdominal adiposity by probiotics (*Lactobacillus gasseri* SBT2055) in adults with obese tendencies in a randomized controlled trial. *Eur J Clin Nutr*. 2010 Mar 10. [Epub ahead of print]
31. Allen SJ, Okoko B, Martinez E, Gregorio G, Dans LF. Probiotics for treating infectious diarrhoea. *Cochrane Database Syst Rev* 2004: CD003048.
32. Huang JS, Bousvaros A, Lee JW, Diaz A, Davidson EJ. Efficacy of probiotic use in acute diarrhea in children: a meta-analysis. *Dig Dis Sci* 2002; 47:2625–2634.
33. McFarland LV. Meta-analysis of probiotics for the prevention of traveler's diarrhea. *Travel Med Infect Dis* 2007, 5:97-105.
34. Culligan EP, Hill C and Sleator RD. Probiotics and gastrointestinal disease: successes, problems and future prospects. *Gut Pathogens* 2009, 1:19.
35. Szajewska H, Kotowska M, Mrukowicz JZ, Armanska M, Mikolajczyk W. Efficacy of *Lactobacillus GG* in prevention of nosocomial diarrhea in infants. *J Pediatr* 2001, 138:361-365.
36. Dubey AP, Rajeshwari K, Chakravarty A, Famularo G. Use of VSL[sharp]3 in the treatment of rotavirus diarrhea in children: preliminary results. *J Clin Gastroenterol* 2008, 42(Suppl 3 Pt 1):S126-129.

37. Rembacken BJ, Snelling AM, Hawkey PM, Chalmers DM, Axon AT. Non-pathogenic *Escherichia coli* versus mesalazine for the treatment of ulcerative colitis: a randomised trial. *Lancet*. 1999;354:635– 639.
38. Kato K, Mizuno S, Umesaki Y, Ishii Y, Sugitani M, Imaoka A, et al. Randomized placebo-controlled trial assessing the effect of bifidobacteria-fermented milk on active ulcerative colitis. *Aliment Pharmacol Ther*. 2004;20:1133–1141.
39. Bibiloni R, Fedorak RN, Tannock GW, Madsen KL, Gionchetti P, Campieri M, et al. VSL#3 probiotic-mixture induces remission in patients with active ulcerative colitis. *Am J Gastroenterol*. 2005;100:1539 –1546.
40. Vanderpool C, Yan F, Polk DB. Mechanisms of probiotic action: implications for therapeutic applications in inflammatory bowel diseases. *Inflamm Bowel Dis* 2008; 14:1585–1596.
41. Kruis W, Frick P, Pokrotnieks J, Lukás M, Fixa B, Kascák M, et al. Maintaining remission of ulcerative colitis with the probiotic *Escherichia coli* Nissle 1917 is as effective as with standard mesalazine. *Gut*. 2004;53:1617–1623.
42. Zocco MA, Dal Verme LZ, Cremonini F, Piscaglia AC, Nista EC, Candelli M, et al. Efficacy of *Lactobacillus GG* in maintaining remission of ulcerative colitis. *Aliment Pharmacol Ther*. 2006;23:1567–1574.

43. Gionchetti P, Rizzello F, Helwig U, Venturi A, Lammers KM, Brigidi P, et al. Prophylaxis of pouchitis onset with probiotic therapy: a double-blind, placebo-controlled trial. *Gastroenterology*. 2003;124:1202–1209.
44. Mimura T, Rizzello F, Helwig U, Poggioli G, Schreiber S, Talbot IC, et al. Once daily high dose probiotic therapy (VSL#3) for maintaining remission in recurrent or refractory pouchitis. *Gut*. 2004;53:108–114.
45. Lammers KM, Vergopoulos A, Babel N, Gionchetti P, Rizzello F, Morselli C, et al. Probiotic therapy in the prevention of pouchitis onset: decreased interleukin-1beta, interleukin-8, and interferon-gamma gene expression. *Inflamm Bowel Dis*. 2005;11:447–454.
46. Pronio A, Montesani C, Butteroni C, Vecchione S, Mumolo G, Vestri A, et al. Probiotic administration in patients with ileal pouch-anal anastomosis for ulcerative colitis is associated with expansion of mucosal regulatory cells. *Inflamm Bowel Dis*. 2008;14:662–668.
47. Kuisma J, Mentula S, Jarvinen H, Kahri A, Saxelin M, Farkkila M. Effect of *Lactobacillus rhamnosus* GG on ileal pouch inflammation and microbial flora. *Aliment Pharmacol Ther*. 2003;17:509–515.
48. Schultz M, Timmer A, Herfarth HH, Sartor RB, Vanderhoof JA, Rath HC.

- Lactobacillus GG in inducing and maintaining remission of Crohn's disease. *BMC Gastroenterol.* 2004;4:5.
49. Bousvaros A, Guandalini S, Baldassano RN, Botelho C, Evans J, Ferry GD, et al. A randomized, double-blind trial of Lactobacillus GG versus placebo in addition to standard maintenance therapy for children with Crohn's disease. *Inflamm Bowel Dis.* 2005;11:833– 839.
50. Marteau P, Lémann M, Seksik P, Laharie D, Colombel JF, Bouhnik Y, et al. Ineffectiveness of Lactobacillus johnsonii LA1 for prophylaxis of postoperative recurrence in Crohn's disease: a randomised, double blind, placebo controlled GETAID trial. *Gut.* 2006;55:842– 847.
51. Prantera C, Scribano ML, Falasco G, Andreoli A, Luzi C. Ineffectiveness of probiotics in preventing recurrence after curative resection for Crohn's disease: a randomised controlled trial with Lactobacillus GG. *Gut.* 2002;51:405–409.
52. Parkes GC, Brostoff J, Whelan K, Sanderson JD. Gastrointestinal microbiota in irritable bowel syndrome: their role in its pathogenesis and treatment. *Am J Gastroenterol* 2008; 103:1557–1567.
53. Quigley EM. The efficacy of probiotics in IBS. *J Clin Gastroenterol* 2008; 42 (Suppl 2):S85–S90.

54. Zeng J, Li YQ, Zuo XL, Zhen YB, Yang J, Liu CH. Clinical trial: effect of active lactic acid bacteria on mucosal barrier function in patients with diarrhoea-predominant irritable bowel syndrome. *Aliment Pharmacol Ther* 2008; 28:994–1002.
55. Moayyedi P, Ford AC, Talley NJ, Cremonini F, Foxx-Orenstein AE, Brandt LJ, et al. The efficacy of probiotics in the therapy of irritable bowel syndrome: a systematic review. *Gut*. 2010; 59(3):325-32.
56. Nikfar S, Rahimi R, Rahimi F, Derakhshani S, Abdollahi M, et al. Efficacy of probiotics in irritable bowel syndrome: a meta-analysis of randomized, controlled trials. *Dis Colon Rectum* 2008;51:1775–1780.
57. McFarland LV, Dublin S. Meta-analysis of probiotics for the treatment of irritable bowel syndrome. *World J Gastroenterol* 2008;14:2650–2661.
58. Hoveyda N, Heneghan C, Mahtani KR, Perera R, Roberts N, Glasziou P. A systematic review and metaanalysis: probiotics in the treatment of irritable bowel syndrome. *BMC Gastroenterol* 2009; 9:15.
59. Bin-Nun A, Bromiker R, Wilschanski M, Kaplan M, Rudensky B, Caplan M, et al. Oral Probiotics Prevent Necrotizing Enterocolitis in Very Low Birth Weight Neonates. *The Journal of Pediatrics* 2005, 147:192-196.
60. Lin HC, Su BH, Chen AC, Lin TW, Tsai CH, Yeh TF, et al. Oral probiotics reduce the

incidence and severity of necrotizing enterocolitis in very low birth weight infants.

Pediatrics 2005, 115:1-4.

61. Caplan MS. Probiotic and prebiotic supplementation for the prevention of neonatal necrotizing enterocolitis. *J Perinatol* 2009; 29 (Suppl 2):S2-S6.

62. Deshpande G, Rao S, Patole S. Probiotics for prevention of necrotizing enterocolitis in preterm neonates with very low birthweight: a systematic review of randomised controlled trials. *Lancet* 2007; 369:1614-1620.

63. Alfaleh K, Anabrees J, Bassler D. Probiotics reduce the risk of necrotizing enterocolitis in preterm infants: a meta-analysis. *Neonatology* 2009; 97: 93-99.

64. D'Souza AL, Rajkumar C, Cooke J, Bulpitt CJ. Probiotics in prevention of antibiotic associated diarrhoea: meta-analysis. *BMJ* 2002, 324:1361.

65. McFarland LV. Meta-analysis of probiotics for the prevention of antibiotic associated diarrhea and the treatment of *Clostridium difficile* disease. *Am J Gastroenterol* 2006, 101:812-822.

66. Johnston BC, Supina AL, Ospina M, Vohra S. Probiotics for the prevention of pediatric antibiotic-associated diarrhea. *Cochrane Database Syst Rev* 2007:CD004827.

67. Canducci F, Armuzzi A, Cremonini F, Cammarota G, Bartolozzi F, Pola P, et al. A

- lyophilized and inactivated culture of *Lactobacillus acidophilus* increases *Helicobacter pylori* eradication rates. *Aliment Pharmacol Ther.* 2000;14:1625–9.
68. Sheu B, Wu J, Lo C, Wu H, Chen J, Lin Y, et al. Impact of supplement with *Lactobacillus* and *Bifidobacterium*-containing yogurt on triple therapy for *Helicobacter pylori* eradication. *Aliment Pharmacol Ther.* 2002;16:1669–75.
69. Sykora J, Valeckova K, Amlerova J, Siala K, Dedek P, Watkins S, et al. Effects of a specially designed fermented milk product containing probiotic *Lactobacillus casei* DN-114 001 and the eradication of *H. pylori* in children: a prospective randomized, double-blind study. *J Clin Gastroenterol.* 2005; 39:692–8.
70. Midolo PD, Lambert JR, Hull R, Luo F, Grayson ML. In vitro inhibition of *Helicobacter pylori* NCTC 11637 by organic acids and lactic acid bacteria. *J Appl Bacteriol.* 1995;79:475–9.
71. Aiba Y, Suzuki N, Kabir AM, Takagi A, Koga Y. Lactic acid-mediated suppression of *Helicobacter pylori* by the oral administration of *Lactobacillus salivarius* as a probiotic in a gnotobiotic murine model. *Am J Gastroenterol.* 1998;93:2097–101.
72. Coconnier MH, Lievin V, Hemery E, Servin AL. Antagonistic activity against *Helicobacter* infection in vitro and in vivo by the human *Lactobacillus acidophilus* strain LB. *Appl Environ Microbiol.* 1998; 64:4573–80.

73. Michetti P, Dorta G, Wiesel PH, Brassart D, Verdu E, Herranz M, et al. Effect of whey-based culture supernatant of *Lactobacillus acidophilus* (johnsonii) La1 on *Helicobacter pylori* infection in humans. *Digestion*. 1999;60:203–9.
74. Lorca GL, Wadstrom T, Valdez GF, Ljungh A. *Lactobacillus acidophilus* autolysins inhibit *Helicobacter pylori* in vitro. *Curr Microbiol*. 2001; 42:39–44.
75. Nam H, Ha M, Bae O, Lee Y. Effect of *Weissella confusa* strain PL9001 on the adherence and growth of *Helicobacter pylori*. *Appl Environ Microbiol*. 2002;68:4642–5.
76. Pinchuk IV, Bressollier P, Verneuil B, Fenet B, Sorokulova IB, Megraud F, et al. In vitro anti-*Helicobacter pylori* activity of the probiotic strain *Bacillus subtilis* 3 is due to secretion of antibiotics. *Antimicrob Agents Chemother*. 2001;45:3156–61.
77. Cats A, Kuipers EJ, Bosschaert MA, Pot RG, Vandenbroucke-Grauls CM, Kusters JG. Effect of frequent consumption of a *Lactobacillus casei*-containing milk drink in *Helicobacter pylori*-colonized subjects. *Aliment Pharmacol Ther*. 2003;17:429–35.
78. Kim TS, Hur JW, Yu MA, Cheigh CI, Kim KN, Hwang JK, et al. Antagonism of *Helicobacter pylori* by bacteriocins of lactic acid bacteria. *J Food Prot*. 2003;66:3–12.
79. Sgouras D, Maragkoudakis P, Petraki K, Martinez-Gonzalez B, Eriotou E,

- Michopoulos S, et al. In vitro and in vivo inhibition of *Helicobacter pylori* by *Lactobacillus casei* strain Shirota. *Appl Environ Microbiol.* 2004;70: 518–26.
80. Canducci F, Armuzzi A, Cremonini F, Cammarota G, Bartolozzi F, Pola P, et al. A lyophilized and inactivated culture of *Lactobacillus acidophilus* increases *Helicobacter pylori* eradication rates. *Aliment Pharmacol Ther.* 2000;14:1625–9.
81. De Francesco V, Stoppino V, Sgarro C, Faleo D. *Lactobacillus acidophilus* administration added to omeprazole/amoxicillin-based double therapy in *Helicobacter pylori* eradication. *Dig Liver Dis.* 2000; 32:746–7.
82. Armuzzi A, Cremonini F, Bartolozzi F, Canducci F, Candelli M, Ojetti V, et al. The effect of oral administration of *Lactobacillus GG* on antibiotic-associated gastrointestinal side-effects during *Helicobacter pylori* eradication therapy. *Aliment Pharmacol Ther.* 2001;15:163–9.
83. Cremonini F, Di Caro S, Covino M, Armuzzi A, Gabrielli M, Santarelli L, et al. Effect of different probiotic preparations on anti-*Helicobacter pylori* therapy-related side effects: a parallel group, triple blind, placebo-controlled study. *Am J Gastroenterol.* 2002;97:2744–9.
84. Sheu B, Wu J, Lo C, Wu H, Chen J, Lin Y, et al. Impact of supplement with *Lactobacillus* and *Bifidobacterium*-containing yogurt on triple therapy for

- Helicobacter pylori* eradication. *Aliment Pharmacol Ther.* 2002;16:1669–75.
85. Myllyluoma E, Veijola L, Ahlroos T, Tynkkynen S, Kankuri E, Vapaatalo H, et al. Probiotic supplementation improves tolerance to *Helicobacter pylori* eradication therapy—a placebo-controlled, double-blind, randomized pilot study. *Aliment Pharmacol Ther.* 2005;21:1263–72.
86. Sykora J, Valeckova K, Amlerova J, Siala K, Dedek P, Watkins S, et al. Effects of a specially designed fermented milk product containing probiotic *Lactobacillus casei* DN-114 001 and the eradication of *H. pylori* in children: a prospective randomized, double-blind study. *J Clin Gastroenterol.* 2005; 39:692–8.
87. Tursi A, Brandimarte G, Giorgetti GM, Modeo ME. Effect of *Lactobacillus casei* supplementation on the effectiveness and tolerability of a new second-line 10-day quadruple therapy after failure of a first attempt to cure *Helicobacter pylori* infection. *Med Sci Monit.* 2005; 10:CR662–6.
88. Anukam KC, Osazuwa EO, Osadolor HB, Bruce AW, Reid G. Yogurt containing probiotic *Lactobacillus rhamnosus* GR-1 and *L. reuteri* RC-14 helps resolve moderate diarrhea and increases CD4 count in HIV/AIDS patients. *J Clin Gastroenterol* 2008; 42:239–243.
89. Wenner M. A cultured response to HIV. *Nat Med* 2009; 15:594–597.

90. Martinez RC, Franceschini SA, Patta MC, Quintana SM, Gomes BC, De Martinis EC, et al. Improved cure of bacterial vaginosis with single dose of tinidazole (2 g), *Lactobacillus rhamnosus* GR-1, and *Lactobacillus reuteri* RC-14: a randomized, double-blind, placebo-controlled trial. *Can J Microbiol* 2009; 55:133–138.
91. Uehara S, Monden K, Nomoto K, Seno Y, Kariyama R, Kumon H. A pilot study evaluating the safety and effectiveness of *Lactobacillus* vaginal suppositories in patients with recurrent urinary tract infection. *Int J Antimicrob Agents* 2006, 28(Suppl 1):S30-34.
92. Kalliomaki M, Salminen S, Poussa T, Arvilommi H, Isolauri E. Probiotics and prevention of atopic disease: 4-year follow-up of a randomised placebo-controlled trial. *Lancet* 2003, 361:1869-1871.
93. Cotter PD, Hill C, Ross RP. Bacteriocins: developing innate immunity for food. *Nat Rev*. 2005;3:777–788.
94. Servin AL. Antagonistic activities of lactobacilli and bifidobacteria against microbial pathogens. *FEMS Microbiol Rev*. 2004;28:405– 440.
95. Lawton EM, Ross RP, Hill C, Cotter PD. Two-peptide lantibiotics: a medical perspective. *Mini Rev Med Chem*. 2007;7:1236 –1247.
96. Morgan SM, O'connor PM, Cotter PD, Ross RP, Hill C. Sequential actions of the two

- component peptides of the lantibiotic lactacin 3147 explain its antimicrobial activity at nanomolar concentrations. *Antimicrob Agents Chemother.* 2005;49:2606–2611.
97. Makarova K, Slesarev A, Wolf Y, Sorokin A, Mirkin B, Koonin E, et al. Comparative genomics of the lactic acid bacteria. *Proc Natl Acad Sci U S A.* 2006;103:15611–15616.
98. Chaillou S, Champomier-Vergès MC, Cornet M, Crutz-Le Coq AM, Dudez AM, Martin V, et al. The complete genome sequence of the meat-borne lactic acid bacterium *Lactobacillus sakei* 23K. *Nat Biotechnol.* 2005;23:1527–1533.
99. Altermann E, Russell WM, Azcarate-Peril MA, Barrangou R, Buck BL, McAuliffe O, et al. Complete genome sequence of the probiotic lactic acid bacterium *Lactobacillus acidophilus* NCFM. *Proc Natl Acad Sci U S A.* 2005;102:3906–3912.
100. Pridmore RD, Berger B, Desiere F, Vilanova D, Barretto C, Pittet AC, et al. The genome sequence of the probiotic intestinal bacterium *Lactobacillus johnsonii* NCC 533. *Proc Natl Acad Sci U S A.* 2004;101:2512–2517.
101. Makras L, Triantafyllou V, Fayol-Messaoudi D, Adriany T, Zoumpoulou G, Tsakalidou E, et al. Kinetic analysis of the antibacterial activity of probiotic lactobacilli towards *Salmonella enterica* serovar Typhimurium reveals a role for lactic acid and other inhibitory compounds. *Res Microbiol.* 2006;157:241–247.

102. De Keersmaecker SC, Verhoeven TL, Desair J, Marchal K, Vanderleyden J, Nagy I, et al. Strong antimicrobial activity of *Lactobacillus rhamnosus* GG against *Salmonella typhimurium* is due to accumulation of lactic acid. *FEMS Microbiol Lett.* 2006;259:89–96.
103. Collado MC, Meriluoto J, Salminen S. Role of commercial probiotic strains against human pathogen adhesion to intestinal mucus. *Lett Appl Microbiol.* 2007;45:454–460.
104. Candela M, Seibold G, Vitali B, Lachenmaier S, Eikmanns BJ, Brigidi P, et al. Real-time PCR quantification of bacterial adhesion to Caco-2 cells: competition between bifidobacteria and enteropathogens. *Res Microbiol.* 2005;156:887–895.
105. Roselli M, Finamore A, Britti MS, Mengheri E, et al. Probiotic bacteria *Bifidobacterium animalis* MB5 and *Lactobacillus rhamnosus* GG protect intestinal Caco-2 cells from the inflammation-associated response induced by enterotoxigenic *Escherichia coli* K88. *Br J Nutr.* 2006;95:1177–1184.
106. Sherman PM, Johnson-Henry KC, Yeung HP, Ngo PS, Goulet J, Tompkins TA. Probiotics reduce enterohemorrhagic *Escherichia coli* O157:H7- and enteropathogenic *E. coli* O127:H6-induced changes in polarized T84 epithelial cell monolayers by reducing bacterial adhesion and cytoskeletal rearrangements. *Infect*

- Immun. 2005;73:5183–5188.
107. Mukai T, Kaneko S, Matsumoto M, Ohori H. Binding of *Bifidobacterium bifidum* and *Lactobacillus reuteri* to the carbohydrate moieties of intestinal glycolipids recognized by peanut agglutinin. *Int J Food Microbiol.* 2004;90:357–362.
108. Sun J, Hu XL, Le GW, Shi YH. Factors involved in binding of *Lactobacillus plantarum* Lp6 to rat small intestinal mucus. *Lett Appl Microbiol.* 2007;44:79–85.
109. Tallon R, Arias S, Bressollier P, Urdaci MC. Strain- and matrix-dependent adhesion of *Lactobacillus plantarum* is mediated by proteinaceous bacterial compounds. *J Appl Microbiol.* 2007;102:442–451.
110. Paton AW, Jennings MP, Morona R, Wang H, Focareta A, Roddam LF, et al. Recombinant probiotics for treatment and prevention of enterotoxigenic *Escherichia coli* diarrhea. *Gastroenterology.* 2005;128:1219–1228.
111. Focareta A, Paton JC, Morona R, Cook J, Paton AW. A recombinant probiotic for treatment and prevention of cholera. *Gastroenterology* 2006;130:1688–1695.
112. Ropeleski MJ, Tang J, Walsh-Reitz MM, Musch MW, Larson RA, Chang EB. Interleukin-11-induced heat shock protein 25 confers intestinal epithelial-specific cytoprotection from oxidant stress. *Gastroenterology* 2003; 124: 1358–1368.
113. Arvans DL, Vavricka SR, Ren H, Musch MW, Kang L, Rocha FG, et al. Luminal

- bacterial flora determines physiological expression of intestinal epithelial cytoprotective heat shock proteins 25 and 72. *Am J Physiol Gastrointest Liver Physiol* 2004; 288: G696–G704.
114. Hu S, Ciancio MJ, Lahav M, Fujiya M, Lichtenstein L, Anant S, et al. Translational inhibition of colonic epithelial heat shock proteins by IFN-gamma and TNF-alpha in intestinal inflammation. *Gastroenterology*. 2007;133:1893–1904.
115. Ludwig D, Stahl M, Ibrahim ET, Wenzel BE, Drabicki D, Wecke A, et al. Enhanced intestinal expression of heat shock protein 70 in patients with inflammatory bowel diseases. *Dig Dis Sci*. 1999;44:1440–1447.
116. Tao Y, Drabik KA, Waypa TS, Musch MW, Alverdy JC, Schneewind O, et al. Soluble factors from *Lactobacillus GG* activate MAPKs and induce cytoprotective heat shock proteins in intestinal epithelial cells. *Am J Physiol Cell Physiol*. 2006;290:C1018–1030.
117. Fujiya M, Musch MW, Nakagawa Y, Hu S, Alverdy J, Kohgo Y, et al. The *Bacillus subtilis* quorum-sensing molecule CSF contributes to intestinal homeostasis via OCTN2, a host cell membrane transporter. *Cell Host Microbe*. 2007 Jun 14;1(4):299-308.
118. Zyrek AA, Cichon C, Helms S, Enders C, Sonnenborn U, Schmidt MA. Molecular

- mechanisms underlying the probiotic effects of *Escherichia coli* Nissle 1917 involve ZO-2 and PKC ζ redistribution resulting in tight junction and epithelial barrier repair. *Cell Microbiol.* 2007;9:804–816.
119. Seth A, Yan F, Polk DB, Rao RK. Probiotics ameliorate hydrogen peroxide-induced epithelial barrier disruption by PKC and MAP kinase-dependent mechanism. *Am J Physiol Gastrointest Liver Physiol.* 2008;294:G1060–G1069.
120. Parassol N, Freitas M, Thoreux K, Dalmaso G, Bourdet-Sicard R, Rampal P. *Lactobacillus casei* DN-114 001 inhibits the increase in paracellular permeability of enteropathogenic *Escherichia coli*-infected T84 cells. *Res Microbiol.* 2005;156:256–262.
121. Otte JM, Podolsky DK. Functional modulation of enterocytes by gram-positive and gram-negative microorganisms. *Am J Physiol Gastrointest Liver Physiol.* 2004;286:G613–G626.
122. Resta-Lenert S, Barrett KE. Probiotics and commensals reverse TNF α - and IFN- γ -induced dysfunction in human intestinal epithelial cells. *Gastroenterology.* 2006;130:731–746.
123. Bakker-Zierikzee AM, Tol EA, Kroes H, Alles MS, Kok FJ, Bindels JG. Faecal SIgA secretion in infants fed on pre- or probiotic infant formula. *Pediatr Allergy Immunol.*

2006;17:134–140.

124. He F, Morita H, Kubota A, Ouwehand AC, Hosoda M, Hiramatsu M, et al. Effect of orally administered non-viable *Lactobacillus* cells on murine humoral immune responses. *Microbiol Immunol.* 2005;49:993–997.
125. Wehkamp J, Harder J, Wehkamp K, Wehkamp-von Meissner B, Schlee M, Enders C, et al. NF- κ B and AP-1-mediated induction of human beta defensin-2 in intestinal epithelial cells by *Escherichia coli* Nissle 1917: a novel effect of a probiotic bacterium. *Infect Immun.* 2004;72:5750–5758.
126. Schlee M, Harder J, Köten B, Stange EF, Wehkamp J, Fellermann K. Probiotic lactobacilli and VSL#3 induce enterocyte beta-defensin 2. *Clin Exp Immunol.* 2008;151:528–535.
127. Pena JA, Versalovic J. *Lactobacillus rhamnosus* GG decreases TNF- α production in lipopolysaccharide-activated murine macrophages by a contact-independent mechanism. *Cell Microbiol.* 2003;5:277–285.
128. Sturm A, Rilling K, Baumgart DC, Gargas K, Abou-Ghazalé T, Raupach B, et al. *Escherichia coli* Nissle 1917 distinctively modulates T-cell cycling and expansion via toll-like receptor 2 signaling. *Infect Immun.* 2005;73:1452–1465.
129. Matsumoto S, Hara T, Hori T, Mitsuyama K, Nagaoka M, Tomiyasu N, et al.

- Probiotic *Lactobacillus*-induced improvement in murine chronic inflammatory bowel disease is associated with the down-regulation of pro-inflammatory cytokines in lamina propria mononuclear cells. *Clin Exp Immunol.* 2005;140:417– 426.
130. Kim SO, Sheikh HI, Ha SD, Martins A, Reid G, et al. G-CSF-mediated inhibition of JNK is a key mechanism for *Lactobacillus rhamnosus*-induced suppression of TNF production in macrophages. *Cell Microbiol.* 2006;8:1958 –1971.
131. Drakes M, Blanchard T, Czinn S. Bacterial probiotic modulation of dendritic cells. *Infect Immun.* 2004;72:3299 –3309.
132. Hart AL, Lammers K, Brigidi P, Vitali B, Rizzello F, Gionchetti P, et al. Modulation of human dendritic cell phenotype and function by probiotic bacteria. *Gut.* 2004;53:1602–1609.
133. Smits HH, Engering A, van der Kleij D, de Jong EC, Schipper K, van Capel TM, et al. Selective probiotic bacteria induce IL-10-producing regulatory T cells in vitro by modulating dendritic cell function through dendritic cell-specific intercellular adhesion molecule 3-grabbing nonintegrin. *J Allergy Clin Immunol.* 2005;115:1260 –1267.
134. Mazmanian SK, Liu CH, Tzianabos AO, Kasper DL, et al. An immunomodulatory molecule of symbiotic bacteria directs maturation of the host immune system. *Cell.*

2005;122:107–118.

135. Ma D, Forsythe P, Bienenstock J. Live *Lactobacillus reuteri* is essential for the inhibitory effect on tumor necrosis factor alpha-induced interleukin-8 expression.

Infect Immun. 2004;72:5308–5314.

136. O'Hara AM, O'Regan P, Fanning A, O'Mahony C, Macsharry J, Lyons A, et al.

Functional modulation of human intestinal epithelial cell responses by *Bifidobacterium infantis* and *Lactobacillus salivarius*. *Immunology.*

2006;118:202–215.

137. Zhang L, Li N, Caicedo R, Neu J, et al. Alive and dead *Lactobacillus rhamnosus* GG

decrease tumor necrosis factor-alpha-induced interleukin-8 production in Caco-2 cells. *J Nutr.* 2005;135:1752–1756.

138. Petrof EO, Claud EC, Sun J, Abramova T, Guo Y, Waypa TS, et al. Bacteria-free

solution derived from *Lactobacillus plantarum* inhibits multiple NF-kappaB pathways and inhibits proteasome function. *Inflamm Bowel Dis.* 2009

Oct;15(10):1537-47.

139. Kelly D, Campbell JI, King TP, Grant G, Jansson EA, Coutts AG, et al. Commensal

anaerobic gut bacteria attenuate inflammation by regulating nuclear-cytoplasmic shuttling of PPAR- γ and RelA. *Nat Immunol.* 2004;5:104–112.

140. Are A, Aronsson L, Wang S, Greicius G, Lee YK, Gustafsson JA, et al. Enterococcus faecalis from newborn babies regulate endogenous PPAR- γ activity and IL-10 levels in colonic epithelial cells. *Proc Natl Acad Sci U S A*. 2008;105:1943–1948.
141. Nenci A, Becker C, Wullaert A, Gareus R, van Loo G, Danese S, et al. Epithelial NEMO links innate immunity to chronic intestinal inflammation. *Nature*. 2007;446:557–561.
142. Grabig A, Paclik D, Guzy C, Dankof A, Baumgart DC, Erckenbrecht J, et al. Escherichia coli strain Nissle 1917 ameliorates experimental colitis via toll-like receptor 2- and toll-like receptor 4-dependent pathways. *Infect Immun*. 2006;74:4075–4082.
143. Rachmilewitz D, Katakura K, Karmeli F, Hayashi T, Reinus C, Rudensky B, et al. Toll-like receptor 9 signaling mediates the anti-inflammatory effects of probiotics in murine experimental colitis. *Gastroenterology*. 2004;126:520–528.
144. Peltekova VD, Wintle RF, Rubin LA, Amos CI, Huang Q, Gu X, et al. Functional variants of OCTN cation transporter genes are associated with Crohn disease. *Nat Genet*. 2004 May;36(5):471-5.
145. Singh J, Rivenson A, Tomita M, Shimamura S, Ishibashi N, Reddy BS. Bifidobacterium longum, a lactic acid-producing intestinal bacterium inhibits colon

- cancer and modulates the intermediate biomarkers of colon carcinogenesis. *Carcinogenesis*, 1997, 18, 833–841.
- 146.Reddy BS, Hamid R, Rao CV. Effect of dietary oligofructose and inulin on colonic preneoplastic aberrant crypt foci inhibition. *Carcinogenesis*, 1997, 18, 1371–1374.
- 147.Sekine K, Ohta J, Onishi M, Tatsuki T, Shimokawa Y, Toida T, et al. Analysis of antitumor properties of effector cells stimulated with a cell wall preparation (WPG) of *Bifidobacterium infantis*. *Biological and Pharmaceutical Bulletin*. 1995, 18, 148–153.
- 148.Okawa T, Niibe H, Arai T, Sekiba K, Noda K, Takeuchi S, et al. Effect of LC9018 combined with radiation therapy on carcinoma of the uterine cervix. *Cancer* 1993, 72, 1949–1954.
- 149.Zhang XB, Ohta Y. Binding of mutagens by fractions of the cell wall skeleton of lactic acid bacteria on mutagens. *Journal of Dairy Science* 1991, 74, 1477–1481.
- 150.Orrhage K, Sillerstrom E, Gustaffson JA, Nord CE, Rafter J. Binding of mutagenic heterocyclic amines by intestinal and lactic acid bacteria. *Mutation Research* 1994, 311, 239–248.
- 151.Zhang XB, Ohta Y. Microorganisms in the gastrointestinal tract of the rat prevent absorption of the mutagen-carcinogen 3-amino-1,4-dimethyl-5H-pyrido [4,3-b]

indole. *Canadian Journal of Microbiology* 1993, 39, 841–845.

152. Baricault L, Denariaz G, Hourii JJ, Bouley C, Sapin C, Trugnan G. Use of HT-29, a cultured human colon cancer cell line, to study the effect of fermented milks on colon cancer cell growth and differentiation. *Carcinogenesis* **1995**, 16, 245 – 252.

Figure legends

Figure 1 Mechanisms of the probiotic functions

Probiotics can exert physiological functions through the inhibition of pathogenic bacteria growth and the adherence of pathogenic bacteria or their toxin to intestinal epithelia, the induction of cytoprotective heat shock proteins, the enhancement of the intestinal barrier function, and modulations of the host immune responses.

Figure 2 Bacterial components or soluble factors released by bacteria are sensed by the intestinal epithelia in some manner (proposal).

Probiotics release bacterial components and effectors which are possibly sensed by TLRs or transported by cell membrane transporters such as OCTN2. Activated downstream of TLRs led to the modulation of host immune responses, while probiotics-derived effectors imported by cell membrane transporters are thought to activate some cell signaling pathway and induce Hsps, thus leading the protection of the intestinal epithelia.

CSF, competence and sporulation factor; LPS, lipopolysaccharide; PG, peptidoglycan; OCTN2, novel organic cation transporter 2; TLR, Toll-like receptor; MAPK, mitogen-activated protein kinase; TRAF6, TNF receptor-associated factor 6; Hsp, Heat

shock protein; NF- κ B, nuclear factor- κ B;